

Immunohistochemical demonstration of specific antigens in the human brain fixed in zinc-ethanol-formaldehyde

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Abstract

Tissue fixation is critical for immunohistochemistry. Recently, we developed a zinc-ethanol-formalin fixative (ZEF), and the present study was aimed to assess the applicability of the ZEF for the human brain histology and immunohistochemistry and to evaluate the detectability of different antigens in the human brain fixed with ZEF. In total, 11 antigens were tested, including NeuN, neuron-specific enolase, GFAP, Iba-1, calbindin, calretinin, choline acetyltransferase, glutamic acid decarboxylase (GAD65), tyrosine hydroxylase, synaptophysin, and α -tubulin. The obtained data show that: i) the ZEF has potential for use in general histological practice, where detailed characterization of human brain morphology is needed; ii) the antigens tested are well-preserved in the human brain specimens fixed in the ZEF.

Introduction

Morphological studies of the human brain are commonly conducted on nervous tissue obtained in autopsy. Quality of the obtained material critically depends on post-mortem time and the fixative chosen. Early autopsy is not always available because of ethical considerations and, accordingly, the fixation became the most crucial step in the preparation of the human brain tissue for morphological observation. The fixation is required to prevent the specimens from the undergoing autolysis (the degradation of macromolecules including proteins and nucleic acids) and decomposition. On the other hand, the fixation must preserve the antigenic properties of the proteins making available their immunohistochemical labeling.

Brain specimens fixed in ethanol or neutral buffered formalin have conventionally been used for classic histological examination with various histological stains. However, both fixatives have disadvantages in immunohistochemistry, because formalin as an electrophilic substance forms crosslinks with reactive functional groups in proteins, nucleic acids, and other macromolecules and, thus, the sterically modified epitopes may have compromised immunoreactivity,^{1,3} whereas ethanol does not form crosslinks, but fixes the tissue by precipitation and the ethanol-fixed tissues showed contraction of the cells.⁴ Therefore, the ethanol-, or formalin-fixed human brain specimens demonstrate bad antigenicity, which impedes or even makes it impossible the immunohistochemical investigation of the tissue. Moreover, the neutral buffered formalin was found to be the poorest fixative for maintaining antigen recognition by immunohistochemistry and the ethanol was also bad for all antigens used.^{5,6} Therefore, a search for alternative fixatives, which offers better preservation of the histology and antigenicity of different organs, has intensively been carried out from the 1990s.⁷⁻¹⁵

Recently, encouraging data have been published that zinc-containing fixatives augment the antigenicity of the tissue and improves the intensity of immunolabeling for some antigens.^{4,5,16-22} Based on these data, we have developed a zinc-containing fixative as a mixture of zinc (as a zinc chloride), ethanol, and formalin and used it successfully for histological and immunohistochemical study of the brain and peripheral tissues of the rat and rabbit.^{23,32}

The aim of the present study was to assess the preservation of different antigens in the human brain fixed in the zinc-ethanol-formalin and the detectability of the antigens using immunohistochemistry performed with various antibodies. The antibodies chosen were those that are conventionally used to label the certain cell type (neurons, glia, ependymocytes), or to show the neurotransmitter specificity of the nerve cells and synapses in the brain.

Materials and Methods

Human brain specimens

Brains were obtained at autopsy from neurologically normal individuals (case details are shown in Table 1). The following brain areas were used in the study: precentral gyrus of parietal lobe, the head and body of the caudate nucleus, *substantia nigra*, nucleus ruber, cerebellar cortex, and, additionally, a thoracic sympathetic ganglion of the sympathetic trunk. The study program was positively approved by

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the local Ethics Committee of the Institute of Experimental Medicine, (St. Petersburg, Russia).

Tissue preparations

The autopsy tissue was immediately sectioned to approximately 10x10x10-mm blocks and placed into the zinc-ethanol-formalin fixative (ZEF). The ZEF was prepared as follows: 1 g ZnCl₂ was dissolved in mixture of 90 mL 96% ethanol and 10 mL concentrated (35-39%) formaldehyde (all purchased from Vekton, St. Petersburg, Russia). Two brain specimens (males, 46 and 51 years-old, *post-mortem* interval 96 h) were fixed in 10% neutral buffered formalin (Vekton, St. Petersburg, Russia).

After fixation for 24 h at room temperature the blocks were dehydrated and embedded in paraffin routinely. Sections 5 or 10 μ m thick were cut using rotary or sliding microtome (RM 2125RT and SM 2000R, respectively; Leica, Wetzlar, Germany) and mounted on poly-L-lysine-coated (Polysine™, Menzel-Gläser, Germany) or silane treated

(HistoBond[®], Marienfeld, Germany) glass slides.

Immunohistochemistry

For immunohistochemistry the sections were deparaffinized and rehydrated routinely. The heat-induced antigen retrieval was carried out by incubating the sections in modified citrate buffer, pH 6.1 (S1700, Dako, Glostrup, Denmark) in a conventional steamer, which steam chamber size allows putting in a Hellendahl jar with the slides, for 25 min. The heat-induced antigen retrieval was omitted for choline acetyltransferase, NeuN, and neuron-specific enolase antibodies. The preparations were pretreated with the blocking solution (Protein Block, Spring Bioscience) and then a primary antibody was applied (details in Table 2). To visualize the applied primary antibodies the following reagents were used: for mouse monoclonal antibodies, MACH2 Mouse HRP-Polymer (Biocare Medical, Concord, CA, USA); for rabbit mono- and polyclonal antibodies, Reveal Polyvalent HRP DAB (Spring Bioscience, Pleasanton, CA, USA); for goat antibodies, biotinylated anti-goat antibody (Dako) and streptavidin conjugated with HRP (streptavidin/horse radish peroxidase; Spring Bioscience). Control of the immunocytochemical reaction was performed according to the recommendation of the reagent manufacturers. All the preparations were counterstained with alum hematoxylin or astra blue and were

examined with a Leica DM 750 microscope and photographed with Leica ICC 50 digital camera (Leica) operated by LAS EZ software (ver. 1.8.0, Leica Microsystems, Heerbrugg, Switzerland).

Results

Histological examination

ZEF-fixed brain preparations were initially Nissl-stained with conventional stains (*not*

shown) and showed good preservation of the nervous tissue obtained from all the studied brain areas, and superior staining of neurons, glial, and endothelial cells. Shrinkage of tissues and swelling of cells was rarely observed and the histology of the brain sections examined was judged to be adequate for research and diagnostic purposes.

Immunohistochemical visualization of antigens

Usage of various immunohistochemical markers demonstrated a pronounced

Table 1. Cases and tissue history.

Case no.	Sex	Age (years)	Cause of death	PMI (h)
1	Male	25	Massive blood loss	15
2	Male	30	Massive blood loss	20
3	Male	34	Mechanical asphyxia	24
4	Male	39	Acute severe asthma	24
5	Female	42	Massive blood loss	24
6	Male	46	Acute cardiac failure	24
7	Male	46	Acute cardiac failure	12
8	Female	48	Aortic rupture	21
9	Male	58	Acute cardiac failure	24
10	Male	60	Acute cardiac failure	24
11	Female	78	Colorectal cancer	18
12	Male	89	Colorectal cancer	72

PMI, *post-mortem* interval.

Table 2. Characteristics of the antibodies used and details of their application.

Antigens	Antibodies (species, clonity, manufacturer, catalogue number, dilution)	Duration and temperature of incubation with antibodies	Visualization
Calbindin	Mouse monoclonal (clone CL-300); Abcam, Cambridge, UK; ab9481; 1:100	100 min, 40°C	MACH2
Calretinin	Rabbit polyclonal; Spring Bioscience; E1170; 1:100	30 min, 40°C	HRP-conjugate
Choline acetyltransferase (ChAT)	Goat polyclonal; Merck Millipore, Billerica, MA, USA; AB144P; 1:250	Overnight, 27°C	Anti-Gt-Bio+Str/HRP*
Glutamic acid decarboxylase (GAD65)	Rabbit polyclonal; Spring Bioscience; E3310; 1:100	30 min, 40°C	HRP-conjugate
Glial fibrillary acidic protein (GFAP)	Mouse monoclonal (clone SPM507); Spring Bioscience; E16510; 1:100	60 min, 40°C	MACH2
Iba-1 protein (AIF1)	Goat polyclonal; Abcam; ab5076; 1:200	overnight, 27°C	Anti-Gt-Bio+Str/HRP*
NeuN protein	Mouse monoclonal (clone A60); Chemicon, Temecula, CA, USA; AB377; 1:400	60 min, 40°C	MACH2
Neuron-specific enolase (NSE)	Rabbit polyclonal; Chemicon; AB951; 1:250	30 min, 40°C	HRP-conjugate
Synaptophysin	Rabbit monoclonal (clone SP11); Spring Bioscience; M3110; 1:100	60 min, 40°C	HRP-conjugate
Tyrosine hydroxylase	Rabbit polyclonal; Abcam; ab112; 1:1000 Mouse monoclonal (clone IB5); Leica-Novocastra, Newcastle-upon-Tyne, UK; NCL-TH36011; 1:50	60 min, 40°C 60 min, 40°C	HRP-conjugate MACH2
α -Tubulin	Mouse monoclonal (clone DM-1A); BioGenex, Fremont, CA, USA; MU121-UC; 1:100	60 min, 40°C	MACH2

MACH2, mouse HRP-Polymer (Biocare Medical); HRP-conjugate from Reveal Polyvalent HRP DAB s(Spring Bioscience); *biotinylated anti-goat antibody (Dako) and streptavidin conjugated with HRP (streptavidin/horse radish peroxidase; Spring Bioscience).

immunostaining of cells and/or fibers in all the examined brain areas (Figure 1). Minimal background staining ensured a contrastive and reproducible image of the immunostained tissue. Quality of images was good irrespectively the counterstaining with hematoxylin or astra blue. We failed to find any visible differences in quality of both histological and immunohis-

tochemical preparations in relation to the post-mortem interval of the brain specimens used in this study.

Differential intracellular immunoreactivity was observed, e.g., tyrosine hydroxylase antibodies clearly labeled cytoplasm to greater extent than the nucleus, while the opposite was true for the NeuN. Cell-specific

immunoreactivity was quite evident for the cell type and neurotransmitter markers used: only astroglia-like ramified cells were labeled with GFAP (Figure 1L), only a part of cell population in the area studied were labeled with tyrosine hydroxylase (Figure 1 D,E), and only some large neurons were choline acetyltransferase-immunoreactive in the caudate nucleus (Figure 1C). The used antibodies were effective for all cortical and subcortical areas examined and in preparations obtained from one and the same brain (Figure 2).

Sections of the human brain fixed in 10% neutral buffered formalin showed good preservation of the nervous tissue in histological preparations (*data not shown*). However, results of immunostaining with the same antibodies were clearly worse with exception of GFAP (Figure 3).

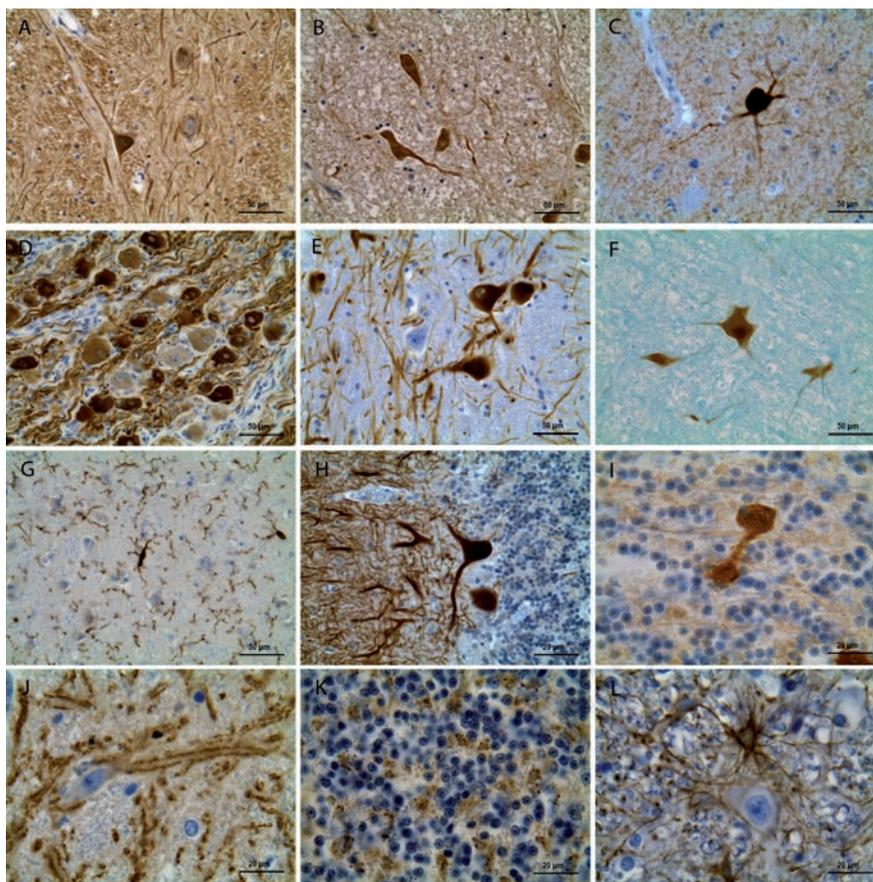


Figure 1. Examples of immunohistochemical staining of the human brain fixed in the ZEF. A) *Substantia nigra, pars reticulata*: α -tubulin in the single neuron with extensively stained perikaryon and processes, many immunoreactive fibers in the surrounding neuropil. B) Nucleus ruber: extensive neuron-specific enolase (NSE) immunoreactivity in the nucleus, perinuclear cytoplasm, and processes of neurons; some bright NSE-immunoreactive fibers and medium-intensity staining of the neuropil are also seen. C) Caudate nucleus (head): choline acetyltransferase (ChAT) immunoreactivity in the multipolar neuron and its processes. D) Thoracic sympathetic ganglion: tyrosine hydroxylase (rabbit polyclonal) immunoreactivity in most of large and small neurons, mainly in the cytoplasm, as well as in bundles of nerve fibers going alongside these neurons. E) *Substantia nigra, pars compacta*: tyrosine hydroxylase (mouse monoclonal) antibody labels many (but not all) nigral neurons. F) Nucleus ruber: NeuN-labeled neurons with highly stained nucleus and the perinuclear cytoplasm. G) Parietal cortex, layer III: Iba-1 distinctly labels multiple thin processes and some perikarya of microgliaocytes. H) Cerebellum: strong calbindin immunostaining in the cytoplasm and dendrites of the Purkinje cells. I) Cerebellum: calretinin immunohistochemistry visualizes the unipolar brush cell, a rare sort of the large cerebellar interneurons in the granular layer.²³ J) *Substantia nigra, pars reticulata*: distinct synaptophysin immunoreactivity in the clusters of synapses alongside the neuronal and fiber plasmalemma. K) Cerebellar cortex: GAD65 immunostaining of the fine fiber nets in the granular layer, which localization coincides with the distribution of the glomeruli, the specific synaptic formations composed by the granular cell dendrites, mossy fibers, and axons of the Golgi cells. L) Nucleus ruber: GFAP distinctly labels the perinuclear cytoplasm and processes of fibrous astrocytes forming large nets in the neuropil. Preparations A-E), G-L) are counterstained with hematoxylin; F) with astra blue.

Discussion

Our assessment of the ZEF for fixation of animal brain for further histological and immunohistochemical studies demonstrated high-quality results for many antibodies.²⁴⁻³³ The study reported here was carried out on post-mortal brains and demonstrated that the ZEF fixation results in superior preservation of the human brain tissue: no tissue shrinkage or distortion of cellular structure was observed, the cellular components retained in their native compartments and the cells were presented with a distinct and detailed microscopic appearance. The tinctorial properties of the ZEF-fixed human brain can be considered excellent: the Nissl-stained preparations produced superior view of all cell types in all the brain areas studied, regardless of whether they were stained with hematoxylin or astra blue. The same good results were obtained after luxol fast blue staining (Klüver-Barrera method) as well (*data not shown*).

Usage of the ZEF-fixed preparations for immunohistochemistry showed that a number of different antigens were present and preserved and that their immunohistochemical visualization was effective. The antigens used included markers of neurons (NeuN and NSE), astro- and microglia (GFAP and Iba-1, respectively), synaptic vesicles (synaptophysin), a protein of microtubules α -tubulin, the neurotransmitter markers choline acetyltransferase, glutamic acid decarboxylase, and tyrosine hydroxylase typical for cholinergic, GABAergic, and catecholaminergic neurons, respectively as well as calcium-binding proteins calbindin and calretinin. The immunohistochemical technique revealed all the used antigens distinctly and with minimal background staining.

Examination of many specimens obtained

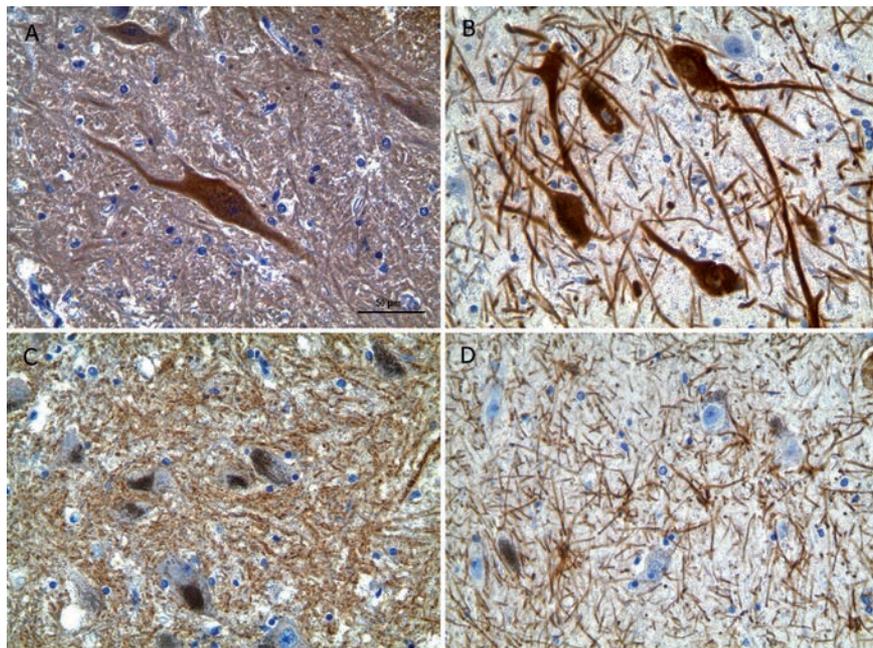


Figure 2. Immunohistochemistry for neuron-specific enolase (NSE), tyrosine hydroxylase (rabbit polyclonal), synaptophysin, and GFAP in the *substantia nigra* of one and the same brain (case No. 4). A) NSE. B) Tyrosine hydroxylase. C) Synaptophysin. D) GFAP. Intensive specific immunostaining of four randomly selected antigens; to be noted the absence of background staining; counterstaining with hematoxylin. Scale bars: 50 µm.

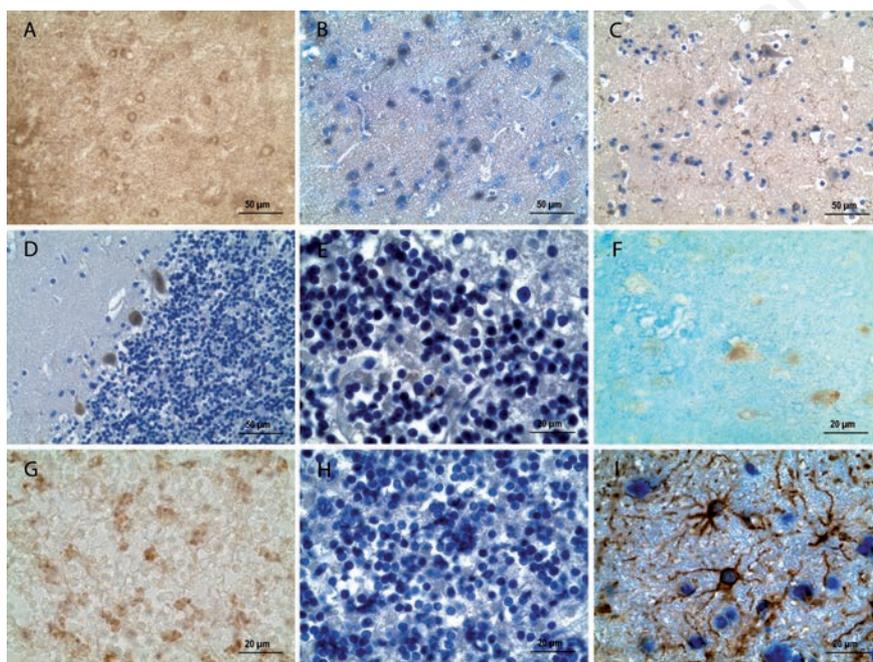


Figure 3. Examples of immunohistochemical staining of the human brain fixed in 10% neutral buffered formalin. A) Parietal cerebral cortex, layer III: α -tubulin, weak immunopositive reaction in neurons virtually indistinguishable from the background. B) Parietal cerebral cortex, layer II-III: NSE, weak immunopositive reaction in some neurons. C) Parietal cerebral cortex, layer II-III: Iba-1, weak immunopositive reaction in microglia. D) Cerebellar cortex: calbindin, weak immunopositive reaction in Purkinje cells. E) Cerebellar cortex: calretinin, negligible immunopositive reaction in cells. F) Parietal cerebral cortex, layer II: NeuN, weak immunopositive reaction in some neurons. G) Cerebellar cortex: synaptophysin, weak immunopositive reaction in glomeruli. H) Cerebellar cortex: GAD65, no immunopositive reaction in cerebellar cells. I) Parietal cerebral cortex, III layer: GFAP, intensive immunopositive reaction in astrocytes. Sections in panels A) and G): without counterstaining; B-E), H), and I): counterstained with hematoxylin; F) with astra blue.

after fixation with ZEF showed greater sensitivity of the immunohistochemical reactions for most of the antigens studied than after use of other fixatives, particularly 10% formalin.³³ Use of fluorescence and confocal laser microscopy methods for animal brain tissue fixed in the ZEF gives images of high quality.^{20,34,35} There was no significant increase in baseline autofluorescence as compared with alcohol and formalin fixation. Of importance is that the ZEF is an inexpensive and easy prepared fixative.

In conclusion, the obtained data indicate that sufficient antigens are preserved in the human brain fixed in the ZEF and that immunohistochemistry technique is applicable and effective for this material. Moreover, the ZEF fixative has potential for use in general histological practice, where detailed characterization of tissue morphology is needed.

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