Astragalus IV ameliorates the dry eye injury in rabbit model via MUC1-ErbB1 pathway

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The therapeutic effects and potential mechanisms of astragaloside IV on a rabbits dry eye model induced by benzalkonium chloride (BAC) was examined. In our study, a BAC-induced dry eye rabbit model was treated with eye drops containing astragaloside IV (5, 10 μM) or solvent four times a day. The clinical evaluations, such as tear break-up time (BUT) and Schirmer tear test (STT), were performed on days 0, 7, 14, 21, and 28. On day 28, the cornea and bulbar conjunctiva tissues (left eye and right eye) were collected with histology, and immunofluorescent staining conducted. The levels of MUC1 and ErbB1 in the corneas were determined by real-time quantitative PCR (qRT-PCR) and the proteins levels of MUC1 and ErbB1 were detected by Western blot. It was demonstrated that both astragaloside IV (5, 10 μM) treatments resulted in an increased STT and BUT on days 7, 14, 21 and 28. Additionally, the astragaloside IV (5, 10 μM)-treated group showed increasing PAS-positive goblet cells than model group (0 μM). Moreover, the MUC1 in model group (0 μM) was decreased, while the expression of MUC1 in astragaloside IV (5, 10 μM) group was increased. Furthermore, astragaloside IV had a protective effect on BAC-induced rabbits’ dry eye and demonstrated clinical improvements, which indicated that astragaloside IV served as a potential protective agent in the clinical treatment of dry eye.

Key words: Astragaloside IV; dry eye model; MUC1; ErbB1.

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Contributions: LC, SM, carried out the experiments; WC, designed the study and edited the manuscript. All authors read and approved the final manuscript and agreed to be accountable for all aspects of the work.

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Availability of data and materials: All data are available from the corresponding authors on reasonable request.

Ethical Approval: The experiment was conducted in accordance with the National Institutes of Health Laboratory Animal Care and Use Guidelines (National Research Council, 1996, USA).
Introduction

Dry eye disease has a complex etiology and it is an eye disease affecting the physical and mental health of 5-50% of the global population. Patients with dry eye disease often suffer ocular discomfort which leads to a lower quality of life. Aqueous deficient dry eye and evaporative dry eye are the most common causes of dry eyes. Aqueous deficiency affects the lacrimal gland, while evaporative dry eye is more associated with the eyelid (e.g., meibomian gland dysfunction and blink abnormalities) or the ocular surface (e.g., related to mucin deficiency or contact lens wear).

Astragaloside IV is a single compound with CAS number 83207-58-3 and molecular weight 784.9702, high-purity drug extracted from *Astragalus membranaceus*, and is also known as “super Astragalus polysaccharide”. Astragaloside IV enhances the body’s immunity and improves the body’s disease resistance. Moreover, astragaloside IV has been reported to inhibit migration and invasion of lung cancer cells A549 by regulating PKC-α-ERK1/2-NF-κB. In addition, the anti-inflammatory effect of astragaloside IV is achieved by NF-κB regulating inflammatory factors and adhesion molecule expression. The gene product of MUC1 transcription is cleaved after translation into two isolated proteins, which are bound together by non-covalent interactions. MUC1 is mainly found in cornea, conjunctival epithelial cells and tear fluid. Most O-glycosylation sites of MUC1 appear in a highly polymorphic core region with a variable number of tandem repeats. MUC1 can also inhibit tear evaporation. The absence and glycosylation of mucin can lead to the lack and denaturation of glycoloyx mucin barrier, which leads to the change of tear film on ocular surface and dry eye. The family of transmembrane tyrosine kinases, ErbB receptors are expressed dynamically during mammary gland development, while the dynamic expression is generally related to breast cancer in both humans and rodents. Overexpression of either the receptors or ligands in this family is usually presented at terminal metastatic disease and constantly leads to overall poor patient outcomes. ErbB1 can enhance the interaction between MUC1/CT and β-catenin and promote the entry of MUC1/CT-β-catenin complex into the nucleus. Astragaloside IV is mainly used in the treatment of inflammatory diseases due to the anti-inflammatory effect of AIV which were demonstrated by a reduction in pro-inflammatory cytokine levels via regulation of the IL-1β, IL-6, IL-10, and TNF-α and these proteins expression. In this study, the effect of astragaloside IV on BAC induced dry eye was comprehensively discussed from multiple perspectives. We believe that astragaloside IV may be a potential drug for the treatment of dry eye. This study investigated astragaloside IV’s effect on cultured human corneal epithelial cells in *in vitro* and evaluated whether its application could promote ocular surface health in an *in vivo* dry eye model.

Materials and Methods

Animals

In our research, 16 adult New Zealand White rabbits (male=8, 2.5±0.5 kg) without ocular abnormalities were conducted in accordance with the National Institutes of Health Laboratory Animal Care and Use Guidelines (National Research Council, 1996, USA). The normal control group consisted of four unoperated rabbits, and 12 rabbits were treated with 0.1% benzalkonium chloride in their eyes (BAC, Sigma-Aldrich, St. Louis, MO, USA) for 15 days. Three groups (four rabbits per group), namely model group (0 μM), astragaloside IV (5 μM, Sigma-Aldrich, St. Louis, MO, USA)-treated group, and astragaloside IV (10 μM)-treated group, were randomly made up of the rabbits with dry eye condition.

Tear flow

Schlemmer tear test (STT) was performed before and after treatment with astragaloside IV at 0, 7, 14, 21 and 28 days, the length of wetting was measured with a ruler with a gradient of 0.5 mm.

Break-up time (BUT)

One μL of 0.1% liquid sodium fluorescein dropping into the conjunctival sac resulted in three blinks, which was recorded in seconds. Observe under a slit lamp microscope with cobalt blue light (CKX41; Olympus, Tokyo, Japan), until the first dryness appears on the tear film spot. The time was recorded from the eyelid opening to the appearance of the first dry spot and was measured three times in succession. The average value was taken and recorded.

Microscopy

Twenty-eight days later, all rabbits were killed by sodium pentobarbital overdose. As the cornea and bulbar conjunctiva tissue of both eyes were removed, aldehyde fixative was used to immerse them for light microscopy, followed by toluidine blue and basic fuchsin stained 1-μm-thick sections after preparation.

qRT-PCR

The TRIzol reagent (Invitrogen, Carlsbad, CA, USA) was adopted to extract the total RNA of the corneas and a reverse transcription kit (RR047A; Takara, Shiga, Japan) was used to synthesize cDNA. Through using a SYBR Premix Ex Taq Kit (RR420A; Takara), real-time quantitative PCR was conducted based on a StepOne Real-Time PCR System (Applied Biosystems, Alameda, CA, USA) with its primer sequences summarized in Table 1.

Cell lines and treatments

Human corneal epithelial cells (HCECs) belonged to a human SV40 immortalized corneal epithelial cell line (CRL-11135, HCE-2; ATCC, Manassas, VA, USA) between passages 25 and 30. They were cultured in a medium containing 10% fetal bovine serum and 10 ng/mL human epidermal growth factor. Later, 1 μg/mL of BCA was adopted to treat the immortalized HCECs to prepare a dry eye corneal epithelial cell injury model. The cells with dry eye condition randomly made up model group, astragaloside IV (5 μM)-treated group, and astragaloside IV (10 μM)-treated group.

Determination of MUC1 level

Human ELISA Development Kit (R & D System, Inc., Minneapolis, MN, USA) was used to determine MUC1 level in the

<table>
<thead>
<tr>
<th>Table 1. The primer sequences.</th>
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<tr>
<td><strong>Gene</strong></td>
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<tr>
<td>ACTB</td>
</tr>
<tr>
<td>Forward</td>
</tr>
<tr>
<td>Reverse</td>
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<tr>
<td>MUC1</td>
</tr>
<tr>
<td>Forward</td>
</tr>
<tr>
<td>Reverse</td>
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culture medium according to the instructions. The culture medium was collected and centrifuged. Then, the supernatant was collected. The concentrations of MUC1 in the culture medium was calculated from a standard curve.

**HCECs transfection**

GenePharma (Shanghai, China) synthesized si-RNA and si-MUC1. According to the manufacturer’s instructions, si-RNA and si-MUC1 (50 nM) were transfected into HCECs by Lipofectamine 2000 (Invitrogen). After 2 days of growth, the original medium was discarded, and RPMI-1640 culture medium (Gibco, Gaithersburg, MD, USA) was selected to the transfected HCECs.

**Western blotting**

Firstly, corneal protein was extracted and the concentration of total protein of cell extract was measured with a protein detection kit. The membranes were incubated overnight at 4°C with rabbit anti-MUC1 antibody (1:1000; Cell Signaling Technology, Danvers, MA, USA), rabbit anti-ErbB1 antibody (1:1000; Cell Signaling Technology), and horseradish peroxidase (HRP)-conjugated anti-β-actin antibody (1:20,000; Sigma-Aldrich) as a loading control. The membranes were washed three times and incubated with HRP-conjugated goat anti-rabbit IgG (1:10,000; Sigma-Aldrich) for 1 h.

**Hematoxylin and Eosin (HE) staining**

Fresh tissues were fixed with 4% formaldehyde, dehydrated and embedded, and then sliced. The paraffin sections were dewaxed with xylene and different concentrations of ethanol, and then put into hematoxylin dyeing solution for 8 min. After washing with water, the paraffin sections were dewaxed with xylene and different concentrations of ethanol, and then put into hematoxylin dyeing solution for 8 min. Then the paraffin sections were dehydrated with ethanol and xylene and sealed with neutral resin.

**Immunofluorescence**

The tissue samples were embedded in paraffin and sectioned using a microtome (LeicaRM2235, Wentzler, Germany). Afterwards, the sections were incubated with an anti-MUC1 antibody (ab104978,1:500; Abcam, Cambridge, UK) overnight at 4°C; then, the sections were incubated with a Cy3-labeled fluorescent secondary antibodies (1:500; Servicebio, Wuhan, China) for 50 min at room temperature. The nuclei were stained with DAPI. The staining was observed by an inverted phase contrast microscope (Olympus) equipped for fluorescence microscopy (DAPI was excited at 330-380 nm and emitted blue light; Cy3 was excited at 510-560 nm and emits red light). The fluorescence intensity was measured on images using Image Pro Plus 6.0.

**Statistical analysis**

We displayed the data with mean ± SEM using SPSS22.0. The measurements were repeated at least three times and the paired t-test was used for statistical comparison.

### Results

**Effect of the astragaloside IV on the Schirmer test**

The normal and model rabbits were treated with astragaloside IV and STT values were obtained after, as shown in Table 2. In the model (0 μM) group, STT values were markedly decreased as compared to control group from 0 day to 28 days (all p<0.01). The vehicle is unable to make the contralateral eye of model rabbits to produce tears like normal rabbit eyes, there was no increase in the eyes. STT values were increased significantly in astragaloside IV (5 μM) from 14 days to 28 days after instillation compared with the model (0 μM) group (p<0.05, p<0.01, p<0.01). STT values were also increased in astragaloside IV (10 μM) from 7 days to 28 days compared with the model (0 μM) group (all p<0.01). Maximal STT values occurred 28 days after instillation in astragaloside IV-treated (10 μM) eyes (17.87±1.44 mm (mean ± SEM)).

**Effect of the astragaloside IV on the BUT**

As shown in Table 3, there was a general trend of decreasing BUT in the model (0 μM) group as compared to the control group.

### Table 2. The values of Schirmer test in each group treatment with astragaloside IV (mm).

<table>
<thead>
<tr>
<th>Group</th>
<th>Control</th>
<th>0 μM</th>
<th>5 μM</th>
<th>10 μM</th>
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<tbody>
<tr>
<td>0 d</td>
<td>15.87±1.12</td>
<td>8.63±0.72*</td>
<td>8.87±0.22</td>
<td>8.77±0.12</td>
</tr>
<tr>
<td>7 d</td>
<td>17.20±1.10</td>
<td>8.57±0.80*</td>
<td>9.37±0.33</td>
<td>10.89±1.01*</td>
</tr>
<tr>
<td>14 d</td>
<td>17.53±1.03</td>
<td>8.61±0.61*</td>
<td>10.87±1.00*</td>
<td>13.87±1.13*</td>
</tr>
<tr>
<td>21 d</td>
<td>17.88±1.02</td>
<td>8.59±0.54*</td>
<td>12.87±1.10*</td>
<td>15.87±1.02*</td>
</tr>
<tr>
<td>28 d</td>
<td>18.12±1.12</td>
<td>8.62±0.83*</td>
<td>14.87±1.07*</td>
<td>17.87±1.10*</td>
</tr>
</tbody>
</table>

*p<0.01 vs Control group; p<0.05; p<0.01 vs astragaloside IV (0 μM).

### Table 3. Values of BUT in each group treatment with astragaloside IV (s).

<table>
<thead>
<tr>
<th>Group</th>
<th>Control</th>
<th>0 μM</th>
<th>5 μM</th>
<th>10 μM</th>
</tr>
</thead>
<tbody>
<tr>
<td>00 d</td>
<td>15.90±1.01</td>
<td>8.60±0.52*</td>
<td>8.67±0.22</td>
<td>8.57±0.12</td>
</tr>
<tr>
<td>7 d</td>
<td>16.20±1.05</td>
<td>8.47±0.60*</td>
<td>9.27±0.33</td>
<td>10.79±1.01*</td>
</tr>
<tr>
<td>14 d</td>
<td>16.53±1.09</td>
<td>8.71±0.51*</td>
<td>10.97±1.00*</td>
<td>12.99±1.13*</td>
</tr>
<tr>
<td>21 d</td>
<td>16.48±1.15</td>
<td>8.49±0.64*</td>
<td>12.65±1.10*</td>
<td>14.47±1.02*</td>
</tr>
<tr>
<td>28 d</td>
<td>16.62±1.13</td>
<td>8.32±0.73*</td>
<td>14.33±1.07*</td>
<td>16.89±1.10*</td>
</tr>
</tbody>
</table>

*p<0.01 vs Control group; p<0.05; p<0.01 vs astragaloside IV (0 μM).
yet no statistically significant differences were found from 0 day to 28 days. The 5 μM astragaloside IV treatment significantly increased BUT compared to the model (0 μM) group on day 14, 21, 28. There also was an increase in BUT in the 10 μM astragaloside IV-treated group on day 7, 14, 21, 28. Maximal BUT occurred 28 days after instillation in astragaloside IV-treated (10 μM) eyes [16.89±1.10 s (mean ± SEM)].

**HE staining of the effect of astragaloside IV on the cornea and bulbar conjunctiva tissue**

The results of HE staining of cornea and conjunctiva of rabbits showed that the color of the corneal extracellular matrix in dry eye model group was lighter than that in normal group because of dehydration, and the color of corneal matrix returned to normal after adding astragaloside IV. There was no significant difference between 5 μm astragaloside IV and 10 μm astragaloside IV. The number of goblet cells in conjunctival tissue of dry eye model group was significantly less than that of normal group. After adding astragaloside IV, the number of goblet cells in conjunctival tissue increased. There was no significant difference between 5 μm astragaloside IV and 10μm astragaloside IV. The results of HE staining were shown in Figure 1.

**Effect of astragaloside IV on the expression of MUC1 in cornea and bulbar conjunctiva tissue**

In order to find out how astragaloside IV influenced the expression of MUC1 in cornea and bulbar conjunctiva tissue, ELISA, qRT-PCR, western blotting, and immunofluorescence staining for MUC1 in cornea and bulbar conjunctiva tissue were performed. The content of MUC1 in cornea and bulbar conjunctiva tissue of the model (0 μM) group was obviously less than control group (p<0.001). The content of MUC1 in cornea and bulbar conjunctiva tissue of the astragaloside IV (5, 10 μM) gradually increased with the administration concentration, and was significantly higher than that of the model when the administration was 10 μM (p<0.001) (Figure 2A). At the same time, the results showed that mRNA and protein expression of MUC1 decreased in the model (0 μM) group, but increased after the astragaloside IV (5, 10 μM) treatment (Figure 2 B-D). The expression of MUC1 was expressed by the intensity of red fluorescence, it is obvious from Figure 2E, the red fluorescence intensity of 0 μM group was significantly lower than that of the normal group, because there were fewer goblet cells and less MUC1 secretion. After adding astragaloside IV, the red fluorescence intensity increased, indicating that MUC1 secretion was more. There was no difference in the red fluorescence intensity between groups 5 μM and 10 μM group, indicating that the expression of MUC1 in these two groups was similar (Figure 2E).

**Effect of astragaloside IV on the MUC1-ErbB1 activation**

qRT-PCR and Western blotting analysis were respectively used to study the activation of ErbB1 (Figure 3). The study revealed that in the control siRNA groups, BAC induction decreased the mRNA
Figure 2. Effect of astragaloside IV on the expression of MUC1 in the cornea and conjunctival tissue. A) The concentration of MUC1 (ng/mL) was evaluated by ELISA. B) The relative mRNA of MUC1 was evaluated by qRT–PCR. C,D) The relative protein expression of MUC1 was evaluated by Western blot. E) Immunofluorescence detection of the expression of MUC1. Scale bars: 100 μm. ***p<0.001 vs Control group, #p<0.05, ## p<0.01 and ### p<0.001 vs astragaloside IV (0 μM).

Figure 3. Effect of astragaloside IV on the MUC1–ErbB1 activation in the corneas. A) The relative protein expression of MUC1 and ErbB1 was evaluated by qRT-PCR. B,C) The relative protein expression of MUC1 and ErbB1 was evaluated by Western blot. ***p<0.001.
Discussion

Astragaloside IV has a single component, which is clearer than many traditional Chinese medicine compounds and has a clear curative effect. It has good biological activity, anti-bacterial, anti-inflammatory and anti-virus antibacterial, anti-inflammatory and antiviral activity, can improve immunity, and is less toxic to liver and kidney than same Western medicines. However, it has been reported that astragaloside IV has genotoxicity, and the lead content in most astragaloside IV samples exceeds the standard, so it is forbidden for pregnant women.

In the present study, astragaloside IV (5, 10 μM) increased STT and BUT in vivo. The results of STT and BUT were the same as previously reported after topical doxycycline, cyclosporine, or uridine administration. Ocular surface mucin (MUC) is a type of high molecular weight glycosylated protein secreted by goblet cells and non-goblet epithelial cells of conjunctival tissue. There are currently 7 types of MUCs present on the ocular surface, namely MUC1, MUC2, MUC4, MUC5AC, MUC5B, MUC7 and MUC16. Amongst them, MUC1 is the first mucin that was confirmed to be located on the ocular surface, secreted by goblet cells, it is also the main component of the mucin layer in the tear film.

Dry diseases of the ocular surface often lead to a decrease in numbers of goblet cells, which in turn affects MUC synthesis and secretion. It is difficult to restore the stability of the tear film simply by supplementing ocular surface water. Therefore, based on effectively promoting tear secretion, it is possible to increase ocular surface viscosity. The synthesis and expression of protein are important prerequisites for restoring ocular surface health. In BAC-induced dry eye model, the expression of MUC1 was decreased but was increased after astragaloside IV treatments.

According to previous investigations, transmembrane MUCs interacted with members of the ErbB family. A co-immunoprecipitation between ErbB2 and MUC4 (ASGP1 -2) in both the metastatic ascites 13762 rat mammary carcinoma cell line as well as the pregnant rat had already been confirmed by Schroeder et al. The presence of MUC1 in immunoprecipitations is due to heterodimerization complexes between the four ErbB receptors and MUC1. Previous study indicated that all ErbB receptors can be turned into transphosphorylase, which revealed that these complexes formed in this phase. It can be comprehended from experiments that induction of BAC inhibits activation of ErbB1, which was increased by astragaloside IV. As cells transfected with MUC1 siRNA showed a loss of ErbB1 in all groups, the expressions of ErbB1 were markedly decreased as compared to those in control siRNA groups. We suggest that MUC1 may be a key mediator for the stability of ErbB1 and thus has an important effect on ErbB1-mediated dry eye.

In summary, our study showed that astragaloside IV improves the clinical efficacy to a certain extent by increasing the expression of MUC1 in dry eyes induced by 0.1% BAC. However, the safety of astragaloside IV for the treatment of dry eye disease requires further evaluation.

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