Common fragile genes

A. Matsuyama, C.M. Croce, K. Huebner

Kimmel Cancer Center, Thomas Jefferson University, Philadelphia, Penn, USA

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Common chromosome fragile sites show susceptibility to DNA damage, leading to alterations that contribute to cancer development. The cloning and characterization of fragile sites have demonstrated that fragile sites are associated with genes that relate to tumorigenesis. Identification of the basis of instability at fragile sites and the related genes provides an entree to understanding of important aspects of chromosomal instability, a prominent feature of neoplastic genomes. FHIT/FRA3B and WWOX/FRA16D, the most sensitive common fragile genes in the human genome, function as tumor suppressor genes. The common features of these two common fragile genes are summarized, and suggest clues to understanding the relation between genomic instability and tumor biology.

Key words: common fragile site, tumor suppressor gene, tumorigenesis, genomic rearrangement

Correspondence: Ayumi Matsuyama, Kimmel Cancer Center, Thomas Jefferson University, 233 South 10th Street, BLSB Room 1006, Philadelphia, Pennsylvania 19107, USA.

Phone: international +1.215.5034655. Fax: international +1.215.9234498. E-mail: ayumi.matsuyama@mail.jci.tju.edu

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hromosomal fragile sites are non-randomly distributed regions that show gaps, breaks or I rearrangements in metaphase chromosomes under specific culture conditions, generally through inhibited or delayed DNA replication (Sutherland, 1979). Now 119 loci have been identified as chromosomal fragile sites in the Human Genome Database(http://www.ncbi.nlm.nih.gov/LocusLink/. Chromosomal fragile sites are classified into two categories on the basis of their frequency in the population and the chemistry of induction (Table 1). Rare fragile sites are observed in less than 5% of individuals and most (23/31) are induced by folic acid deficiency, whereas common fragile sites are found in all individuals and most (76/88) are induced by aphidicolin, an inhibitor of DNA polymerases (Glover et al., 1984). Unstable repeats are expand at the rare fragile sites and lead to chromosome breakage; for example, folate-sensitive FRA11B is associated with CGG triplets repeats (Jones et al., 1995), and BrdU-induced FRA10B and distamycin A-induced FRA16B with AT-rich minisatellite repeats (Hewett et al., 1998; Yu et al., 1997). The CGG repeats lead to unusual secondary DNA structures, such as hairpins (Gacy et al., 1995), slipped strand (S)-DNA (Pearson et al., 1998), or quadruplex DNA (Fry and Loeb, 1994), these structures tend to inhibit or delay DNA replication in vitro and in vivo (Usdin and Woodford, 1995; Samadashwily et al., 1997). AT-rich minisatellite repeats can also form hairpin structures (Handt et al., 2000).

Common fragile sites

On the other hand, several common fragile sites have been cloned and characterized: FRA3B (Ohta et al., 1996; Inoue et al., 1997; Mimori et al., 1999), FRA6E (Cesari et al., 2003; Denison et al., 2003), FRA6F (Morelli et al., 2002), FRA7G (Huang et al., 1999; Tatarelli et al., 2000), FRA7H (Mishmar et al., 1998), FRA16D (Mangelsdorf et

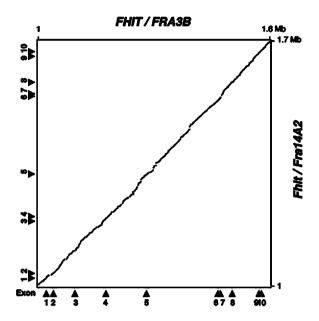


Figure 1. Comparison of human and murine orthologous sequences. The PipMaker program (http://blo.cse.psu.edu/plpmaker/, Schwartz et al., 2000) was applied for the large-scale comparison between human FHIT and mouse Fhit sequences, that computes alignments of similar regions in two DNA sequences. The horizontal and vertical axes represent the nucleotide number of human and mouse sequences, respectively, and the locations of exons in FHIT and Fhit are also provided as arrowheads.

al., 2000; Paige et al., 2000; Ried et al., 2000), and FRAXB (Arlt et al., 2002) (Figure 1). It is now known that these sites are actually huge regions, encompassing megabases, with proneness to forming gaps or breaks, and that sequences of these sites are rich in adenosine and thymidine with many sequence features, such as long interspersed nuclear element 1 (LINE1), high flexibility regions (HFRs), and matrix association regions (MARs), that are sensitive to genomic rearrangements (Inoue et al., 1997; Mimori et al., 1999; Mishmar et al., 1998; Tatarelli et al., 2000; Ried et al., 2000; Shiraishi et al., 2001; Matsuyama et al., 2003). Common fragile sites are induced also by caffeine and alcohol, and environmental carcinogens, such as in cigarette smoke, can enhance their expression (Yunis and Soreng, 1984; Kuwano and Kajii, 1987; Chen et al., 1989; Ban et al., 1995). Fragile sites are recombinogenic and sensitive to agents that inhibit DNA replication. At fragile sites, DNA replication occurs late in the cell cycle, so accumulation of incomplete replication leads to gaps/breaks in metaphase chromosome (Laird et al., 1987). Studies have revealed that the fragile sequences must be responsible for late/delayed

DNA replication (Le Beau et al., 1998; Wang et al., 1999; Hellman et al., 2000), however, no specific sequences that could perturb replication have been identified. A recent study reported that the replication checkpoint kinase ATR is critical for genome stability at fragile sites (Casper et al., 2002). Fragility of common fragile regions occurs spontaneously in ATR deficient cells. Thus, stalling of DNA replication forks that escape ATR checkpoint could lead to persistence of single-stranded regions through G2 and mitosis, appearing as gaps in mitotic chromosomes. Zlotorynski et al (2003) reported that aphidicolin-induced common fragile sites contain flexible sequences, composed of interrupted runs of AT-dinucleotides, which can affect DNA replication by forming secondary structures, and these sequences are similar to the AT-rich minisatellite repeats in rare fragile sites, FRA16B and FRA10B. Rare and common fragile sites, thus, may both depend upon flexible sequences for their fragility. We have determined that the human and mouse fragile histidine triad (FHIT/Fhit) genes, encompassing the FRA3B/Fra14A2 common fragile regions, respectively, have more HFRs in their sequences than the flanking human and mouse protein tyrosine receptor phosphatase (PTPRG/Ptprg) genes (Matsuyama et al., 2003). The results suggest that AT-rich flexible sequences might play an important role in the fragility mechanism in the mammalian genome. If fragility is based on specific sequences, then the specific hallmark sequences of fragile regions, the HFRs, must be responsible for the late/delayed replication phenomenon, as well as the occurrence of singlestrandedness that can escape the ATR DNA damage checkpoint detection.

Fragile sites have been reported also in mouse, rat, hamster, cow, cat, and dog (Djalali et al., 1987; Lin et al., 1984; Smeets and van de Klundert, 1990; Stone et al., 1991; Vitetta et al., 1991; Robinson and Elder, 1987). They appear to be inherent and universal structures of the mammalian genome, and specific fragile regions seem to be conserved across species (Yunis and Soreng, 1984). We already reported that human and mouse orthologous common fragile sites, FRA3B and Fra14A2, are highly conserved (Shiraishi et al., 2001). Sequence comparison revealed that human and mouse orthologous genes, FHIT and Fhit, are more highly conserved through evolution than flanking PTPRG/Ptprg genes (Figure 1, Matsuyama et al.,

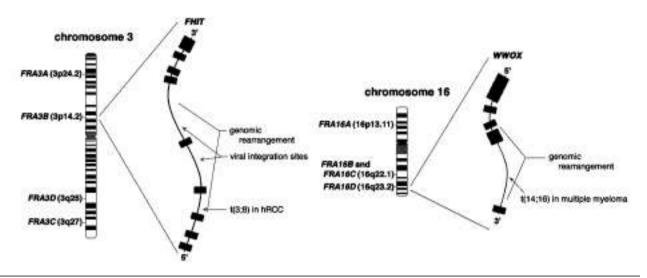


Figure 2. The FHIT and WWOX loci. The FHIT gene encompasses FRA3B, at 3p14.2 on chromosome 3, and the WWOX gene spans 16q23.2 on chromosome 16. FRA3A, FRA3C, FRA3D, and FRA16C are also common fragile sites, and FRA16A and FRA16B are rare ones. Gray boxes represent the location of exons in the sequences.

2003). The human PTPRG gene is located about 1.5 Mbp centromeric to FHIT on chromosome 3; and the mouse ortholog, Ptprg, is located 1.7 Mbp centromeric to Fhit on chromosome 14. The results indicate that not only common fragile sites but also the fragile genes encompassing common fragile sites are recombinogenic.

Chromosome locations of many common fragile sites coincide with locations of cancer breakpoints and/or cancer-associated genes, and the hypothesis that they play an important role in chromosomal instability in cancer development was proposed (Yunis and Soreng, 1984). This hypothesis has produced many studies of recombiningenicity involving common fragile sites, reporting observations of sister chromatid exchanges, translocations, deletions, viral integrations, intrachromosomal gene amplifications, at or near fragile sites in cancer (Glover and Stein, 1988; Popescu et al., 1990; Coquelle et al., 1997; Mishmar et al., 1998). Several genes at an encompassing common fragile sites have been shown to or suggested to function as tumor suppressor genes. Two genes, a tumor suppressor and candidate suppressor associated with the most active common fragile sites, FHIT and WWOX are reviewed below.

FHIT/FRA3B

FRA3B at chromosomal band 3p14.2 is the most active common fragile site in the human genome, and chromosomal abnormalities in this region are

observed in a wide variety of human malignant diseases (reviewed in Huebner et al., 1998; Huebner and Croce, 2001, 2003). FRA3B exhibits hallmarks of fragile site recombinigenicity, such as the translocation, t(3;8)(p14.2;q24), breakpoint at 3p14.2 in familial clear cell renal carcinoma (Cohen et al., 1979), plasmid integration sites (Rassool et al., 1991) and a papilloma virus integration site (Wilke et al., 1996) (Figure 2). The fragile histidine triad (FHIT) gene was found to span FRA3B (Ohta et al., 1996). The FHIT locus spans over 1.5 Mb pairs (Mbp) of human genome, including FRA3B, but the mRNA is only 1.1 kb in size and consists of 10 small exons; exons 5 to 9 encode the Fhit protein (Ohta et al., 1996; Zimonjic et al., 1997) (Figure 2). According to the sequence and features of FRA3B; most cancer associated rearrangements thus far defined occur within introns 3, 4, and 5 of the FHIT gene (Inoue et al., 1997; Mimori et al., 1999), and not only the orthologous fragile sites, human FRA3B and mouse Fra14A2, but also the orthologous genes, FHIT and Fhit, are highly conserved over evolution (Shiraishi et al., 2001; Matsuyama et al., 2003). The region from exon 3 through 6 has been considered the epicenter of fragility for FRA3B and thus this region is referred to as FRA3B and Fra14A2.

FHIT is frequently deleted in cancers and cancer cell lines, such as lung, digestive tract, kidney, breast, liver, and pancreatic cancers, and Fhit protein is absent or reduced in most cancers (reviewed

Table 1. Classification of human chromosomal fragile sites and features.

Classification (n)	Common fragile sites (88)	Rare fragile sites (31) observed in less than 5% of individuals	
Frequency in the popular	tion observed in all individuals		
Inducing chemistry (n)	aphidicolin (76) 5- bromodeoxyuridine (BrdU) (7) 5-azacytidine (4) unclassified (1)	folic acid deficiency (23) distamycin A (5) BrdU (2) unclassified (1)	
Feature of sequence	common feature is still unknown (AT-rich flexible sequence ?) n,	unstable repeat expansion (AT-rich minisatellite repeat, CGG triplets repeat,) number of identified human loci.	

 $\label{thm:common fragile sites related with malignancies.} \label{thm:common fragile sites related with malignancies.}$

Locus	Location	Related genes	Alteration in malignancies
FRA3B	3p14.2	FHIT	deletion in a wide variety of cancers FHIT product is reduced/absent in most cancers translocation in renal cancer
FRA6E	6q26	Parkin	LOH in ovarian and breast cancer duplication or deletion of exons
FRA6F	6q21	19 known genes are related	deletion in several types of leukemias and solid tumors
FRA7G	7q31.2	CAV1, CAV2, TES	LOH in ovarian, breast and prostate cancer
FRA16D	16q23.3~24.1	L WWOX/WOX1/FOR	LOH in liver, ovarian, breast, esophageal, prostate, and lung cancer deletion in ovarian and liver cancer translocations in multiple myeloma
FRAXB	Хр22.31	STS, GS1, TLR5a	deletion in esophageal cancer cells LOH, loss of heterozygosity

in Huebner et al., 1998; Huebner and Croce, 2001, 2003; Richards, 2001). The mouse Fhit ortholog also encompasses a common fragile site, Fra14A2 on murine chromosome 14 (Glover et al., 1998), and sustains homozygous deletions in murine cancer cell lines (Pekarsky et al., 1998). FHIT expression is lost/reduced at an early stage, both in esophageal cancer, adenocarcinoma and squamous cell carcinoma, cancers strongly influenced by environmental carcinogens, such as in tobacco smoke and alcohol consumption (Michael et al., 1997; Mori et al., 2000). Thus, loss of FHIT function is involved in carcinogenesis at an early stage of cancer progression. Exogeneous FHIT can suppress tumor growth (Siprashvili et al., 1997), and exogenous Fhit protein expression induces apoptosis, directly or indirectly, in cancer cells (Ji et al., 1999; Sard et al., 1999). Furthermore, Fhit knockout mice are more susceptible to tumor formation (Fong et al., 2000; Zanesi et al., 2001) and delivery of FHIT in viral vectors prevents and reverses cancer development (Dumon et al., 2001; Ishii et al., 2001; Ishii et al., 2003). Thus, Fhit functions as a tumor suppressor, consistently with the idea that fragile sites may harbor genes that, when altered, contribute to cancer development.

WWOX/FRA16D

Human WW domain containing oxidoreductase (WWOX, also known as WOX1 or FOR) gene was identified from chromosome region 16q23.3-24.1 at the aphidicolin-sensitive common fragile site FRA16D (Bednarek et al., 2000; Ried et al., 2000). At that time FRA16D had been partially

characterized, and, within the region, homozygous deletion in gastric, ovarian, colon cancer, and loss of heterozygosity (LOH) in prostate and breast cancer had been reported (Mangelsdorf et al., 2000; Paige et al., 2000). The murine orthologous Wwox gene also encompasses a common fragile site, Fra8E1, and FRA16D/Fra8E1 are highly conserved, like FRA3B/Fra14A2, across species (Krummel et al., 2002). The WWOX gene encompasses a genomic locus > 1 Mb, consists of 9 small exons with 2.3 kb mRNA (Bednarek et al., 2000). The translocation, t(14;16)(q32;q23), breakpoint in multiple myeloma (Chesi et al., 1998; Krummel et al., 2000) is observed in intron 8 (Figure 2). Homozygous deletions have been defined between exons 4-9 of WWOX in some cancers, liver (Yakicier et al., 2001), ovarian (Paige et al., 2001), breast (Driouch et al., 2002), esophageal (Kuroki et al., 2002), and lung cancer (Yendamuri et al., 2003). WWOX mRNA expression has been examined in tumors and cancer cell lines and aberrant products observed in many tumors, including breast (Bednarek et al., 2001), ovary (Paige et al., 2001), esophageal (Kuroki et al., 2002), and lung cancers (Yendamuri et al., 2003). Wwox protein may engage a proapoptotic pathway that inhibits tumor cell growth in vitro and in vivo (Bednarek et al., 2001), perhaps through c-Jun NH2-terminal kinase interaction (Chang et al., 2003). WWOX is involved in stress and apoptotic responses, and the presence of WW domains in the structure of WWOX indicates that it physically interacts with other proteins (Bednarek et al., 200). WWOX protein has been reported to interact with both p53 and JNK1. The

two fragile genes, FHIT and WWOX, have common features; both encompass a genomic locus of more than 1 Mb with an associated fragile region, show frequent altered expression in cancers, and are suspected tumor suppressor genes.

Other common fragile sites, genes, cancers, and future directions

Cancer-related common fragile sites are summarized in Table 2. FRA7G at 7q31.2 is involved in LOH in ovarian, breast and prostate cancers; three genes near this fragile region, the caveolin-1 and -2 (CAV1 and 2), and the TESTIN (TES) genes, have been reported to show altered expression in cancers (Engelman et al., 1998; Huang et al., 1999; Tatarelli et al., 2000). In addition, CAV1 and 2 have been reported as tumor suppressor genes (Hurlstone et al., 1999).

FRAXB, at Xp22.31, spans a large genomic region of approximately 500 kb, three known genes, including GS1 (a gene of unknown function), the microsomal steroid sulfatase locus (STS), and Toll-like receptor 5a (TLR5a), map near or within this fragile site region (Arlt et al., 2002). In this study, multiple deletions in cancer cell lines within four fragile sites, FRAXB, FRA7G, FRA7H and FRA16D, were reported, supporting the hypothesis that common fragile sites and their associated genes are unstable in some cancer cells.

The common fragile site FRA6F, located at 6q21, is an extended region of about 1.2 Mb; deletion in this region has been recognized in several types of leukemia and solid tumors. Transcription mapping of the FRA6F region identified 19 known genes (Morelli et al., 2002).

Recently, in the third most active common fragile site, FRA6E (6q26), Parkin, the gene involved in the pathogenesis of juvenile Parkinson's disease has been identified (Cesari et al., 2003; Denison et al., 2003). Alterations in FRA6E and Parkin are observed in ovarian, breast, lung and other cancers. Thus, Parkin is a large gene located in a large common fragile site, and is also a candidate tumor suppressor gene, like FHIT/FRA3B and WWOX/FRA16D.

To date, many findings have revealed the relationship between common fragile sites, related genes, and cancer (Table 2). Genomic rearrangements in several genes, located in or encompassing common fragile sites, lead to tumorigenesis and tumor progression; thus, these genes can have roles as tumor

suppressors. Further detailed analyses of genes at common chromosome fragile sites will determine whether genes at these sites commonly have roles as fragile tumor suppressor genes. Since fragile genes are likely to be inactivated in parallel in cancer, the characteristics of biological functions of these fragile genes will contribute to a further understanding of the natural history of cancer progression.

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